

hydroxy- $\delta$ -cadinen-15-oate TLC of the mixture of **1a** and **2a** (Et<sub>2</sub>O-petrol, 3:2) gave 70 mg **2a** (*R<sub>f</sub>* 0.70). Known compounds were identified by comparison of the 400 MHz <sup>1</sup>H NMR spectra with those of authentic material and by co-TLC.

**Methyl 2 $\beta$ -hydroxy- $\delta$ -cadinen-15-oate (1a)** Colourless oil, IR  $\nu_{\text{max}}^{\text{CCl}_4}$  cm<sup>-1</sup> 3560 (OH, hydrogen bonded), 1710, 1655 (C=CCO<sub>2</sub>R), MS *m/z* (rel int) 264 [M]<sup>+</sup> (24) (calc for C<sub>16</sub>H<sub>24</sub>O<sub>3</sub> 264.173), 246 [M-H<sub>2</sub>O]<sup>+</sup> (20), 203 [246-CHMe<sub>2</sub>]<sup>+</sup> (80), 187 [246-CO<sub>2</sub>Me]<sup>+</sup> (100), 176 [246-H<sub>2</sub>C=CHCHMe<sub>2</sub>, RDA]<sup>+</sup> (30), <sup>13</sup>C NMR (C<sub>6</sub>D<sub>6</sub>, C-1-C-15) 33.2 t, 65.4 d, 124.0 s, 144.4 d, 41.0 d, 44.0 d, 21.7 t, 34.6 t, 129.6 s, 133.0 s, 27.0 d, 19.1 q, 21.5 q, 15.7 q, 16.7 s, 51.3 q (OMe).

**Preparation of 1a from the acetate** To 23 mg  $\beta$ -acetoxy- $\delta$ -cadinen-15-oic acid in 2 ml MeOH, 0.5 ml 2N KOH was added. After 2 hr the crude acid was esterified with CH<sub>2</sub>N<sub>2</sub>. TLC (Et<sub>2</sub>O-petrol, 4:1) gave 16 mg **1a** (*R<sub>f</sub>* 0.60) identical with the methyl ester obtained from the natural product (<sup>1</sup>H NMR and co-TLC).

**Methyl 2 $\beta$ -methoxy- $\delta$ -cadinen-15-oate (2a)** Colourless oil, IR  $\nu_{\text{max}}^{\text{CCl}_4}$  cm<sup>-1</sup> 1710, 1650 (C=CCO<sub>2</sub>R), MS *m/z* (rel int) 278 [M]<sup>+</sup> (11) (calc for C<sub>17</sub>H<sub>26</sub>O<sub>3</sub> 278.188), 260 [M-H<sub>2</sub>O]<sup>+</sup> (7), 246 [M-MeOH]<sup>+</sup> (51), 235 [M-CHMe<sub>2</sub>]<sup>+</sup> (9), 203 [235-MeOH]<sup>+</sup> (100), 187 [246-CO<sub>2</sub>Me]<sup>+</sup> (91), 176 [246-H<sub>2</sub>C=CHCHMe<sub>2</sub>, RDA]<sup>+</sup> (45), 145 [187-C<sub>3</sub>H<sub>6</sub>]<sup>+</sup> (76).

**Methyl 13-hydroxy-10 $\alpha$ -peroxy-cadina-3,9(14)-dien-15-oate (3a)** Colourless oil, IR  $\nu_{\text{max}}^{\text{CCl}_4}$  cm<sup>-1</sup> 3600 (OH), 1710 (C=CCO<sub>2</sub>R), MS (CI, isobutane) *m/z* (rel int) 297 [M+1]<sup>+</sup> (62) (calc for C<sub>16</sub>H<sub>24</sub>O<sub>5</sub>+1), 279 [297-H<sub>2</sub>O]<sup>+</sup> (100), 263 [297-H<sub>2</sub>O<sub>2</sub>]<sup>+</sup> (57), 247 [279-MeOH]<sup>+</sup> (38), EI 262 [M-H<sub>2</sub>O<sub>2</sub>]<sup>+</sup> (21), 231 [262-OMe]<sup>+</sup> (32), 203 [231-CO]<sup>+</sup> (42), 61 (100),

$$[\alpha]_{24}^{20} = \frac{578}{-10} \frac{546}{-19} \frac{436 \text{ nm}}{-63} \text{CHCl}_3, c = 0.3$$

**Methyl 13-hydroxy-9 $\alpha$ -peroxy-cadina-1(10),3-dien-15-oate (4a)** Colourless oil, IR  $\nu_{\text{max}}^{\text{CCl}_4}$  cm<sup>-1</sup> 3600 (OH), 1720 (C=CCO<sub>2</sub>R), MS (CI, isobutane) *m/z* (rel int) 297 [M+1]<sup>+</sup> (10), (calc for C<sub>16</sub>H<sub>24</sub>O<sub>5</sub>+1), 279 [297-H<sub>2</sub>O]<sup>+</sup> (21), 263 [297-H<sub>2</sub>O<sub>2</sub>]<sup>+</sup> (14), 247 [279-MeOH]<sup>+</sup> (8), 209 [297-C<sub>5</sub>H<sub>12</sub>O]<sup>+</sup> (100), EI 262 [M-H<sub>2</sub>O<sub>2</sub>]<sup>+</sup> (2.5), 230 [262-MeOH]<sup>+</sup> (40), 61 (100).

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## EUDESMANOLIDES FROM ARTEMISIA JUDAICA

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**Key Word Index**—*Artemisia judaica*, Compositae, sesquiterpenes, eudesmanolides

**Abstract**—The aerial parts of *Artemisia judaica* afforded in addition to known compounds a hydroperoxide corresponding to vulgarin as well as an isomer of the latter. The configurations of these lactones have been established by NOE difference spectroscopy.

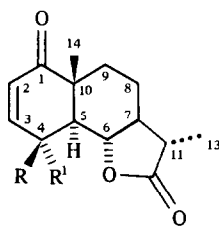
The aerial parts of *Artemisia judaica* L. have been investigated previously [1-3], and we have studied now material collected in Egypt. In addition to tauremisin (= vulgarin, **1**) [4, 5] isolated previously from this species [1, 2], we obtained the isomer **3** and the hydroperoxide **2** and in addition ethyl cinnamate,  $\alpha$ -pinene, chrysanthenone, camphor, piperitone, verbenol and the hydroperoxide **4** isolated so far only from *Artemisia inculta* [6]. The structure of **2** could be deduced from the spectral data of the product obtained by triphenyl phosphine-reduction which were identical with those of **1**. The spectral data of **3** (Table 1) were close to those of **1**. However, the chemical shifts of H-14 and H-15 differed characteristically. NOE

difference spectroscopy with both **1** and **3** clearly indicated the configuration at C-4. While **1** gave clear NOEs between H-14 and H-15 and H-6, the isomer **3** showed NOEs between H-14 and H-6 as well as between H-15 and H-5 and H-3. The configuration of **3** has been assigned previously for a lactone named barrelin [7]. Comparison of the <sup>13</sup>C NMR data, however, show that this lactone most likely is identical with vulgarin though the mp and the optical rotation differ. The published <sup>13</sup>C NMR data of **1** [8] differ only in the chemical shift of C-14 which was erroneously assigned ( $\delta$  22.7 is the value of C-8 and not of C-14 which is 19.7). The <sup>13</sup>C NMR data of **1** and **3** show some clear differences. In particular, the C-15 signal is

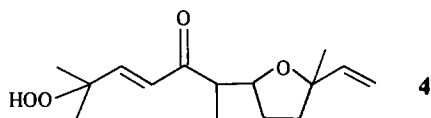
Table 1  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectral data of compounds 1–3 (400 MHz,  $\text{CDCl}_3$ , TMS as int. standard)

	1*	2	3		1	3
H-2	5.90 d	6.00 d	5.87 d	C-1	201.6 s	203.1
H-3	6.61 d	6.74 d	6.50 d	C-2	125.7 d	125.3
H-5	2.43 d	2.82 d	2.02 d	C-3	151.7 d	150.2
H-6	4.16 dd	4.12 dd	4.33 dd	C-4	70.1 s	68.2
H-7	1.67 dddd	1.66 dddd	1.66 dddd	C-5	54.7 d	51.0
H-8 $\alpha$	1.98 dddd	1.98 dddd	2.0 m	C-6	79.6 d	79.2
H-8 $\beta$	1.56 dddd	1.55 m	1.51 m	C-7	52.5 d	52.3
H-9 $\alpha$	1.47 ddd	1.47 m		C-8	22.7 t	22.8
H-9 $\beta$	2.03 ddd	2.04 ddd	2.0 m	C-9	34.3 t	32.5
H-11	2.35 q	2.33 dq	2.33 dq	C-10	46.2 s	45.9
H-13	1.23 d	1.21 d	1.58 d	C-11	40.6 d	40.1
H-14	1.19 s	1.22 s	1.31 s	C-12	178.1 s	179.0
H-15	1.54 s	1.47 d	1.58 s	C-13	12.4 q	12.5
				C-14	19.7 q	20.6
				C-15	23.8 q	31.7

\*Chemical shift slightly varying by concentration

J (Hz) 2, 3 = 10, 5, 6 = 11, 6, 7 = 10, 7, 8 $\alpha$  = 3.5, 7, 8 $\beta$  = 12, 7, 11 = 12, 8 $\alpha$ , 8 $\beta$  = 13, 8 $\alpha$ , 9 $\alpha$  = 3.5, 8 $\alpha$ , 9 $\beta$  = 3.5, 8 $\beta$ , 9 $\alpha$  = 12.5, 8 $\beta$ , 9 $\beta$  = 3, 9 $\alpha$ , 9 $\beta$  = 14, 11, 13 = 7

	1	2	3
R	Me	Me	OH
R <sup>1</sup>	OH	OOH	Me



authentic material. Due to the small amounts 2 and 3 could not be induced to crystallize though they were homogeneous by  $^1\text{H}$  NMR spectroscopy.

4 $\alpha$ -Hydroperoxy-desoxyvulgarin (2) (gum)  $^1\text{H}$  NMR see Table 1. To 5 mg 2 in 0.5 ml  $\text{CDCl}_3$ , 20 mg triphenyl phosphine was added. After 10 min at room temp the  $^1\text{H}$  NMR spectrum had completely changed to that of 1. TLC ( $\text{Et}_2\text{O}$ ) gave 1, identical with authentic vulgarin.

4-Epi-vulgarin (3) (gum) IR  $\nu_{\text{max}}^{\text{CCl}_4}$   $\text{cm}^{-1}$  3600, 3440 (OH), 1790 ( $\gamma$ -lactone), 1690 ( $\text{C}=\text{C}=\text{O}$ ), MS  $m/z$  (rel. int.) 264.136 [ $\text{M}$ ] $^+$  (6) (calc. for  $\text{C}_{15}\text{H}_{20}\text{O}_4$  264.136), 249 [ $\text{M} - \text{Me}$ ] $^+$  (12), 98 (57), 55 (100).

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shifted downfield, most likely due to its quasi-equatorial orientation.

## EXPERIMENTAL

The air-dried aerial parts (220 g, collected from Wadi Hof, Egypt, March 1984) was worked-up in the usual fashion [9]. CC fractions were as follows: 1 (petrol), 2 ( $\text{Et}_2\text{O}$ -petrol, 1:20), 3 ( $\text{Et}_2\text{O}$ -petrol, 1:10), 4 ( $\text{Et}_2\text{O}$ -petrol, 1:4) and 5 ( $\text{Et}_2\text{O}$  and  $\text{Et}_2\text{O}$ -MeOH, 10:1). TLC of fraction 1 (petrol) gave 50 mg  $\alpha$ -pinene and of fraction 2 ( $\text{Et}_2\text{O}$ -petrol, 1:15), 3 mg chrysanthemone, 2 mg camphor, 2 mg ethyl cinnamate and 10 mg piperitone. TLC of fraction 3 ( $\text{Et}_2\text{O}$ -petrol, 1:4) gave 5 mg piperitone and 5 mg verbenol and TLC of fraction 4 ( $\text{Et}_2\text{O}$ -petrol, 9:11) 5 mg 4. TLC of fraction 5 ( $\text{Et}_2\text{O}$ -petrol, 9:1) gave 5 mg 3 ( $R_f$  0.52), 3 mg 1 ( $R_f$  0.40) and 5 mg 2 ( $R_f$  0.35). Known compounds were identified by comparing the spectral data, especially the 400 MHz  $^1\text{H}$  NMR spectra, with those of

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